Pancreatic Cancer Immunology – Setting the Scene

Prof Jen Morton – CRUK Beatson / Glasgow Uni

Disclosures:

Have received research funding from AstraZeneca, UCB Pharma, Redex Pharma, Bioven





Intro - Tumour Immunology 101







Intro – the PDAC immune landscape





Current approaches





Targeting neutrophils/MDSCs

- In mice, neutrophil depletion or blocking trafficking (CXCR2) leads to reduction in tumour or metastatic burden and strongly synergize with PD-1 blockade.
- Mice treated with CCR2i show reduced PDAC tumour burden.
 - But, early trial data for CXCR2i + IO were disappointing
- FOLFIRINOX + CCR2i resulted in reductions in primary tumour burden, leading to downstaging and increased eligibility for surgery.
- Unfortunately, a similar trial of a CCR2i + Gem/nP in metastatic PDAC failed for both safety and efficacy.
- Neutrophil/MDSC blocking strategies cannot be maintained long term due to the critical importance of neutrophils for host defence?





Targeting neutrophils/MDSCs

CellPress

Cancer Cell **Article**

CXCR2 Inhibition Profoundly Suppresses Metastases and Augments Immunotherapy in Pancreatic Ductal Adenocarcinoma

Colin W. Steele,¹ Saadia A. Karim,¹ Joshua D.G. Leach,¹ Peter Bailey,² Rosanna Upstill-Goddard,² Loveena Rishi,² Mona Foth,¹ Sheila Bryson,¹ Karen McDaid,³ Zena Wilson,³ Catherine Eberlein,³ Juliana B. Candido,⁴ Mairi Clarke,⁵ Colin Nixon,¹ John Connelly,¹ Nigel Jamieson,⁶ C. Ross Carter,⁶ Frances Balkwill,⁴ David K. Chang,² T.R. Jeffry Evans.^{1,2} Douglas Strathdee,¹ Andrew V. Biankin,² Robert J.B. Nibbs,⁵ Simon T. Barry,³ Owen J. Sansom,^{1,2,7,*} and Jennifer P. Morton^{1,2,7}

Research **CXCR2-Dependent Accumulation of Tumor-**Associated Neutrophils Regulates T-cell Immunity in Pancreatic Ductal Adenocarcinoma

Timothy Chao¹, Emma E. Furth^{1,2}, and Robert H. Vonderheide^{1,3}

SCIENCE TRANSLATIONAL MEDICINE | RESEARCH ARTICLE

CANCER

Agonism of CD11b reprograms innate immunity to sensitize pancreatic cancer to immunotherapies

Roheena Z. Panni¹, John M. Herndon², Chong Zuo², Samarth Hegde², Graham D. Hogg², Brett L. Knolhoff², Marcus A. Breden², Xiaobo Li³, Varintra E. Krisnawan², Samia Q. Khan³, Julie K. Schwarz^{4,5}, Buck E. Rogers^{4,5}, Ryan C. Fields^{1,4}, William G. Hawkins^{1,4}, Vineet Gupta^{3*}, David G. DeNardo^{2,4,6}*[†]

Pancreas



Research Article

ORIGINAL ARTICLE

Targeting both tumour-associated CXCR2⁺ neutrophils and CCR2⁺ macrophages disrupts myeloid recruitment and improves chemotherapeutic

responses in pancreatic ductal adenocarcinoma

Timothy M Nywening, ^{1,2} Brian A Belt, ^{3,4,5} Darren R Cullinan, ^{1,2} Roheena Z Panni, ^{1,2} Booyeon J Han, ^{3,4,5} Dominic E Sanford, ^{1,2} Ryan C Jacobs, ^{1,2} Jian Ye, ^{3,4,5} Ankit A Patel, ^{3,4,5} William E Gillanders, ^{1,2} Ryan C Fields, ^{1,2} David G DeNardo, ^{2,6,7} William G Hawkins, ^{1,2} Peter Goedegebuure, ^{1,2} David C Linehan^{3,4,5}





Cancer

mmunology

Targeting macrophages

- Macrophages are highly abundant and mostly tumor-promoting (?). Close proximity of alternatively activated TAMs to tumour cell nests correlates with poor prognosis
- Efficacy of macrophage depletion may be impacted by compensatory increases in other myeloid cell populations, however, there are alternative approaches, e.g.:
- Reprogramming macrophages to phagocytose and kill live tumor cells
 - Via agonistic anti-CD40 which can activate the phagocytic program in macrophages.
 - Via NF-κB modulation which can induce lymphotoxin production from T cells to reprogramme macrophages to phagocytose.
 - Via activation of macrophages with the TLR9 ligand CpG which can induce phagocytosis
- Blockade of the 'don't eat me' ligand CD47 on tumor cells





Targeting macrophages – which ones??

- PDAC TAMs originate from bone marrow-derived infiltrating monocytes *and* embryonic tissue resident macrophages (TRMs)
- TRMs exhibit pro-fibrotic activities (distinct from infiltrating monocytes) and only they promote PDAC progression
- Tissue-resident TAMs expand in PDAC via in situ proliferation
- However, during the early stages of tumorigenesis, TRMs could have tumor suppressive properties?
- Should we be specifically targeting the pro-tumorigenic, immunosuppressive signals, rather than the cells themselves?



Immunity Article

Pancreatic

Cancer



Article

Cell

Tissue-Resident Macrophages in Pancreatic Ductal Adenocarcinoma Originate from Embryonic Hematopoiesis and Promote Tumor Progression

Yu Zhu,^{1,2} John M. Herndon,^{1,2} Dorothy K. Sojka,³ Ki-Wook Kim,⁴ Brett L. Knolhoff,^{1,2} Chong Zuo,^{1,2} Darren R. Cullinan, Jingqin Luo,^a Audrey R. Bearden,^{1,2} Kory J. Lavine,¹ Wayne M. Yokoyama,^{3,2} William G. Hawkins,^{6,7} Ryan C. Fields,^{6,7} Gwendalyn J. Randolph,^{1,4} and David G. DeNardo^{1,2,4,5},⁴

Resident Macrophages Cloak Tissue Microlesions to Prevent Neutrophil-Driven Inflammatory Damage

Stefan Uderhardt,^{1,4,*} Andrew J. Martins,² John S. Tsang,² Tim Lämmermann,³ and Ronald N. Germain^{1,*} ¹Umphocyte Biology Section, Laboratory of Immune System Biology, National Institute of Alergy and Infectious Diseases, NIH, Bethesda, MD 20892, USA



Targeting macrophages

Microenvironment and Immunology

Cancer Research

CSF1/CSF1R Blockade Reprograms Tumor-Infiltrating Macrophages and Improves Response to T-cell Checkpoint Immunotherapy in Pancreatic Cancer Models

Yu Zhu^{1,2}, Brett L. Knolhoff^{1,2}, Melissa A. Meyer^{1,2}, Timothy M. Nywening^{3,4}, Brian L. West⁵, Jingqin Luo^{4,6}, Andrea Wang-Gillam¹, S. Peter Goedegebuure^{3,4}, David C. Linehan^{3,4}, and David G. DeNardo^{1,2,4,7}

SCIENCE TRANSLATIONAL MEDICINE | RESEARCH ARTICLE

CANCER

Pancreatic

Cancer

Agonism of CD11b reprograms innate immunity to sensitize pancreatic cancer to immunotherapies

Roheena Z. Panni¹, John M. Herndon², Chong Zuo², Samarth Hegde², Graham D. Hogg², Brett L. Knolhoff², Marcus A. Breden², Xiaobo Li³, Varintra E. Krisnawan², Samia Q. Khan³, Julie K. Schwarz^{4,5}, Buck E. Rogers^{4,5}, Ryan C. Fields^{1,4}, William G. Hawkins^{1,4}, Vineet Gupta^{3*}, David G. DeNardo^{2,4,6*†} immunology

ARTICLES https://doi.org/10.1038/s41590-018-0292-

Metabolic rewiring of macrophages by CpG potentiates clearance of cancer cells and overcomes tumor-expressed CD47—mediated 'don't-eat-me' signal

Mingen Liu¹, Roddy S. O'Connor², Sophie Trefely^{3,4}, Kathleen Graham¹, Nathaniel W. Snyder³ and Gregory L. Beatty 15*



Cell Reports Article

CSF1R⁺ Macrophages Sustain Pancreatic Tumor Growth through T Cell Suppression and Maintenance of Key Gene Programs that Define the Squamous Subtype

Juliana B. Candido,^{1,8} Jennifer P. Morton,^{2,3,8} Peter Bailey,³ Andrew D. Campbell,² Saadia A. Karim,² Thomas Jamieson,² Laura Lapienyte,² Aarthi Gopinathan,⁴ William Clark,² Ewan J. McGhee,² Jun Wang,¹ Monica Escorcio-Correia,¹ Raphael Zollinger,¹ Rozita Roshani,¹ Lisa Drew,⁵ Loveena Rishi,³ Rebecca Arkell,¹ T.R. Jeffry Evans,^{2,3} Colin Nixon,² Duncan I. Jodrell,⁴ Robert W. Wilkinson,⁶ Andrew V. Biankin,³ Simon T. Barry,⁷ Frances R. Balkwill,¹ and Owen J. Sansom^{2,3,9,*}

Pancreas



ORIGINAL ARTICLE

Targeting both tumour-associated CXCR2⁺ neutrophils and CCR2⁺ macrophages disrupts myeloid recruitment and improves chemotherapeutic responses in pancreatic ductal adenocarcinoma

Timothy M Nywening,^{1,2} Brian A Belt,^{3,4,5} Darren R Cullinan,^{1,2} Roheena Z Panni,^{1,2} Booyeon J Han,^{3,4,5} Dominic E Sanford,^{1,2} Ryan C Jacobs,^{1,2} Jian Ye,^{3,4,5} Ankit A Patel,^{3,4,5} William E Gillanders,^{1,2} Ryan C Fields,^{1,2} David G DeNardo,^{2,6,7} William G Hawkins,^{1,2} Peter Goedegebuure,^{1,2} David C Linehan^{3,4,5}

Synergy or not with dual targeting???



Targeting CAFs (to reprogramme the immune TME)

- Blanket CAF targeting has been unsuccessful, however, targeting CAF subsets or their secreted products (e.g. myeloid cell-recruiting chemokines) may be worthwhile, e.g.:
 - anti-TGFβ antibodies can decrease CAFs, reduce myeloid infiltration and relieve CTL suppression.
 - ATRA / PIN1 targeting have demonstrated efficacy in orthotopic mouse models by promoting CAF quiescence and reducing fibrosis and may synergize with chemo + ICB.
 - Blockade of IL-1β (which supports iCAFs to produce IL-6 and and reinforce the immunosuppressive PDAC TME) synergizes with ICB in preclinical models.
- Clinical trials of all of the above concepts are ongoing
- Whether any will be effective remains to be determined....





Targeting CAFs (to reprogramme the immune TME)

Cancer Cell Article

Cancer-Associated Fibroblasts Neutralize the Anti-tumor Effect of CSF1 Receptor Blockade by Inducing PMN-MDSC Infiltration of Tumors

Vinit Kumar,¹ Laxminarasimha Donthireddy,¹ Douglas Marvel,^{1,10} Thomas Condamine,^{1,11} Fang Wang,¹ Sergio Lavilla-Alonso,^{1,12} Ayumi Hashimoto,¹ Prashanthi Vonteddu,¹ Reeti Behera,¹ Marlee A. Goins,⁶ Charles Mulligan,⁶ Brian Nam,⁶ Neil Hockstein,⁶ Fred Denstman,⁶ Shanti Shakamuri,⁶ David W. Speicher,² Ashani T. Weeraratna,¹ Timothy Chao,⁴ Robert H. Vonderheide,⁴ Lucia R. Languino,⁸ Peter Ordentlich,³ Qin Liu,¹ Xiaowei Xu,⁴ Albert Lo,^{5,13} Ellen Puré,⁵ Chunsheng Zhang,⁹ Andrey Loboda,⁹ Manuel A. Sepulveda,⁷ Linda A. Snyder,⁷ and Dmitry I. Gabrilovich^{1,14,*}

CelPress

Article

Type I collagen deletion in αSMA⁺ myofibroblasts augments immune suppression and accelerates progression of pancreatic cancer

Yang Chen,¹ Jiha Kim,¹ Sujuan Yang,¹ Huamin Wang,² Chang-Jiun Wu,³ Hikaru Sugimoto,¹ Valerie S. LeBleu,¹ and Raghu Kalluri^{1,4,*}

Cancer Cell



Article

CellPress

Mesothelial cell-derived antigen-presenting cancer-associated fibroblasts induce expansion of regulatory T cells in pancreatic cancer

Huocong Huang,^{1,2,11,*} Zhaoning Wang,⁷ Yuqing Zhang,^{2,3,9} Rachana N. Pradhan,⁸ Debolina Ganguly,^{2,3} Raghav Chandra,^{1,2} Gilbert Murimwa,^{1,2} Steven Wright,² Xiaowu Gu,^{4,10} Ravikanth Maddipati,^{2,5} Sören Müller,⁸ Shannon J. Turley,⁸ and Rolf A. Brekken^{1,2,3,6,*}

CXCR4 inhibition in human pancreatic and colorectal cancers induces an integrated immune response

Daniele Biasci^{a,b,c,1}, Martin Smoragiewicz^{a,b,1,2},, Claire M. Connell^{a,b,d,1},, Zhikai Wang^{e,1}, Ya Gao^e, James E. D. Thaventhiran^{a,b,c}, Bristi Basu^{d,f}, Lukasz Magiera^{a,b,3}, T. Isaac Johnson^{a,b}, Lisa Bax^d, Aarthi Gopinathan^{a,b}, Christopher Isherwood^{a,b}, Ferdia A. Gallagher^g, Maria Pawula^{a,b}, Irena Hudecova^{a,b}, Davina Gale^{a,b}, Nitzan Rosenfeld^{a,b}, Petros Barmpounakis^h, Elizabeta Cristina Popaⁱ, Rebecca Brais^j, Edmund Godfrey^g, Fraz Mir^k, Frances M. Richards^{a,b}, Douglas T. Fearon^{a,e,i,4,5}, Tobias Janowitz^{a,b,e,I,4,5}, and Duncan I. Jodrell^{a,b,4}



Cancer Cell



Targeting / reprogramming DCs

- Is T cell exhaustion the problem in PDAC or is it a lack of T cell priming?
- In mice, there is a dearth of cross-presenting type 1 conventional dendritic cells (cDC1s) in pancreatic tumours
- Anti-CD40 (agonistic) antibodies can activate DCs and enhance T cell priming.
- FLT3L treatment increases intratumoral cDC1s and restores sensitivity to CD40 agonist antibody and RT in PDAC mouse models.
- Ongoing trials to determine tolerability & efficacy of CD40 agonist + FLT3L in human.
- Other approaches include targeted agents that affect DDR pathways, to activate the STING pathway and IFN production, to promote DC activation.





Targeting / reprogramming DCs



Targeting B cells

- Several high impact papers reported a tumour promoting role in PDAC
- My lab have a lot of evidence that this isn't true.... If nothing else, the conflicting data show that model and context are very important!

IN THE SPOTLIGHT

B Cells Promote Pancreatic Tumorigenesis

Ali Roghanian^{1,2}, Christopher Fraser¹, Marianna Kleyman¹, and Jianzhu Chen¹

Bruton Tyrosine Kinase-Dependent Immune **Cell Cross-talk Drives Pancreas Cancer**

Andrew J. Gunderson¹, Megan M. Kaneda², Takahiro Tsujikawa^{1.3}, Abraham V. Nguyen², Nesrine I. Affara⁴ Brian Ruffell¹, Sara Gorjestani², Shannon M. Liudahl¹, Morgan Truitt⁵, Peter Olson⁵, Grace Kim^{4,6}, Douglas Hanahan⁷, Margaret A. Tempero^{6,8}, Brett Sheppard^{9,10}, Bryan Irving¹¹, Betty Y. Chang¹², Judith A. Varner^{2,13}, and Lisa M. Coussens^{1,10}

IL35-Producing B Cells Promote the **Development of Pancreatic Neoplasia**

Yuliya Pylayeva-Gupta¹, Shipra Das¹, Jesse S. Handler¹, Cristina H. Hajdu², Maryaline Coffre², and Dafna Bar-Sagi¹ Sergei B. Koralov

Hifla Deletion Reveals Pro-Neoplastic Function of B Cells in Pancreatic Neoplasia

Kyoung Eun Lee^{1,2}, Michelle Spata^{1,2}, Lauren J. Bayne^{1,2}, Elizabeth L. Buza³, Amy C. Durham³, David Allman^{2,4}, Robert H. Vonderheide^{1,2}, and M. Celeste Simon^{1,2,5}

Aside: Do we know what tertiary lymphoid structures are doing???





Cancer

Tumour-Intrinsic immune resistance

- mKRAS orchestrates a network of immunosuppression
 - enhances autophagocytosis to modulate cell surface MHC-I
 - regulates expression of CD47 & PD-L1
 - drives GM-CSF & CXCL1 expression

Pancreatic

- downregulates CCL4 to impede DC recruitment
- promotes SHH signalling & expression of COX2, IL-6, pSTAT3, and MMP7 tc
 drive chronic inflammation & fibrosis.
- p53 deficiency promotes M2-like TAM polarization.
- p53^{R172H} mutation drives neutrophil accumulation
- *Cdkn2a, Pten, Lkb1* mutations all associated with IO resistance

Should we consider genetic/transcriptomic subtype when thinking about immune-targeting strategies?



CANCER

RESEARCH

BEATSON INSTITUTE

Targeting T cell function

- ICB (typically targeting PD-1/PD-L1 or CTLA-4) has been been largely unsuccessful, beyond the ~1% of patients with MSI high tumors
- Although T cell numbers can vary, they're rarely located adjacent to tumor cell nests. Thus, efforts have focused on increasing T cell numbers more than targeting function
- But, do we understand T cells well enough???
 - e.g. Are Tregs good or bad???
 - What ICB should we use?? (e.g. TIGIT Pasca di Magliano, Jacks)



Pancreatic

Cancer

Cell Reports

Crosstalk between Regulatory T Cells and Tumor-Associated Dendritic Cells Negates Anti-tumor Immunity in Pancreatic Cancer

Jung-Eun Jang,^{1,2} Cristina H. Hajdu,³ Caroline Liot,² George Miller,^{4,5} Michael L. Dustin,^{1,6,*} and Dafna Bar-Sagi^{2,7,*}

RESEARCH ARTICLE

Regulatory T-cell Depletion Alters the Tumor Microenvironment and Accelerates Pancreatic Carcinogenesis 🛯 🔮

Yaqing Zhang^{1,2}, Jenny Lazarus¹, Nina G. Steele³, Wei Yan¹, Ho-Joon Lee⁴, Zeribe C. Nwosu⁴, Christopher J. Halbrook⁴, Rosa E. Menjivar⁵, Samantha B. Kemp⁶, Veerin R. Sirihorachai⁷, Ashley Velez-Delgado³, Katelyn Donahue⁷, Eileen S. Carpenter⁸, Kristee L. Brown¹, Valerie Irizarry-Negron¹, Anna C. Nevison¹, Alekya Vinta⁹, Michelle A. Anderson⁸, Howard C. Crawford^{2,48}, Costas A. Lyssiotis^{2,48}, Timothy L. Frankel¹, Filip Bednar¹, and Marina Pasca di Magliano^{1,235}



- $\gamma\delta$ T cells
 - $\gamma \delta T$ cells constitute ~40% of infiltrating T cells in PDAC???

Article

$\gamma\delta$ T Cells Support Pancreatic Oncogenesis by Restraining $\alpha\beta$ T Cell Activation

Donnele Daley,^{1,6,7} Constantinos Pantelis Zambirinis,^{1,6,7} Lena Seifert,^{1,6,7} Neha Akkad,¹ Navyatha Mohan,¹ Gregor Werba,¹ Rocky Barilla,¹ Alejandro Torres-Hernandez,¹ Mautin Hundeyin,¹ Vishnu Raj Kumar Mani,¹ Antonina Avanzi,¹ Daniel Tippens,¹ Rajkishen Narayanan,¹ Jung-Eun Jang,^{2,3} Elliot Newman,¹ Venu Gopal Pillarisetty,⁴ Michael Loran Dustin,^{3,5} Dafna Bar-Sagi,² Cristina Hajdu,³ and George Miller^{1,6,8,*}

 Innate lymphoid cells – do they do anything??

Article

ILC2s amplify PD-1 blockade by activating tissue-specific cancer immunity

https://doi.org/10.1038/s41586-020-2015-4 Received: 26 March 2019 Accepted: 31 December 2019

Published online: 19 February 2020

John Alec Moral^{12,3}, Joanne Leung^{12,3,17}, Luis A. Rojas^{12,317}, Jennifer Ruan^{12,3,17}, Julia Zhao^{12,3}, Zachary Sethna^{12,3}, Anita Ramnarain^{12,3}, Billel Gasmi⁴, Murali Gururajan⁵, David Redmond⁶, Gokce Askan⁷, Umesh Bhanot⁷, Ela Elyada^{8,3}, Youngkyu Park^{8,9}, David A. Tuveson^{8,9}, Mithat Gönen¹⁰, Steven D. Leach¹¹, Jedd D. Wolchok^{2,4,12,13,14,15}, Ronald P. DeMatteo¹⁶, Taha Merghoub^{2,4,13,14,15} & Winod P. Balachandran^{12,2,13,13}





Hot topics

• Neoantigen quality, not quantity?

Article

Neoantigen quality predicts immunoediting in survivors of pancreatic cancer

https://doi.org/10.1038/s41586-022-04735-9	Marta Łuksza ^{1,16 🖂} , Zachary M. Sethna ^{2,3,4,16} , Luis A. Rojas ^{3,4,16} , Jayon Lihm², Barbara Bravi ^{5,6} ,
Received: 11 June 2021	Yuval Elhanati ² , Kevin Soares ⁴⁷ , Masataka Amisaki ³⁴ , Anton Dobrin ^{8,9} , David Hoyos ² , Pablo Guasp ^{3,4} , Abderezak Zebboudi ^{3,4} , Rebecca Yu ^{3,4} , Adrienne Kaya Chandra ^{3,4} ,
Accepted: 7 April 2022	Theresa Waters ^{3,4} , Zagaa Odgerel ^{3,4} , Joanne Leung ⁴ , Rajya Kappagantula ^{7,10} ,
Published online: 19 May 2022	Alvin Makohon-Moore ⁷¹⁰ , Amber Johns ¹¹ , Anthony Gill ^{11,12} , Mathieu Gigoux ^{3,13} , ledd Wolchok ^{3,13} Taha Marghouh ^{3,13} Michel Sadelain ^{8,9} Frin Patterson ⁴ Remi Monasson ⁵
Open access	Thierry Mora ⁵ , Aleksandra M. Walczak ⁵ , Simona Cocco ⁵ , Christine Jacobuzio-Donahue ⁷¹⁰ ,
Check for updates	Benjamin D. Greenbaum ^{2,14 Colore} & Vinod P. Balachandran ^{3,4,7,15 Colored}



Tumor Cell-Intrinsic Factors Underlie Heterogeneity of Immune Cell Infiltration and Response to Immunotherapy

Jinyang Li,^{1,14} Katelyn T. Byrne,^{2,5,14,*} Fangxue Yan,¹ Taiji Yamazoe,¹ Zeyu Chen,⁶ Timour Baslan,⁹ Lee P. Richman,¹ Jeffrey H. Lin,¹ Yu H. Sun,¹¹ Andrew J. Rech,^{1,5} David Balli,¹ Ceire A. Hay,² Yogev Sela,¹ Allyson J. Merrell,¹ Shannon M. Liudahl,¹² Naomi Gordon,¹ Robert J. Norgard,¹ Salina Yuan,¹ Sixiang Yu,¹ Timothy Chao,¹ Shuai Ye,¹ T.S. Karin Eisinger-Mathason,¹ Robert B. Faryabi,^{1,6,7} John W. Tobias,⁸ Scott W. Lowe,^{9,10} Lisa M. Coussens,¹² E. John Wherry,^{4,6,6} Robert H. Vonderheide,^{1,2,4,6,6,*} and Ben Z. Stanger^{1,2,3,4,5,13,*}

The NEW ENGLAND JOURNAL of MEDICINE

BRIEF REPORT

Neoantigen T-Cell Receptor Gene Therapy in Pancreatic Cancer

Rom Leidner, M.D., Nelson Sanjuan Silva, B.S., Huayu Huang, M.S., David Sprott, B.S., Chunhong Zheng, Ph.D., Yi-Ping Shih, Ph.D., Amy Leung, B.S., Roxanne Payne, M.N., Kim Sutcliffe, B.S.N., Julie Cramer, M.A., Steven A. Rosenberg, M.D., Ph.D., Bernard A. Fox, Ph.D., Walter J. Urba, M.D., Ph.D., and Eric Tran, Ph.D.

LETTER

doi:10.1038/nature24462

Identification of unique neoantigen qualities in long-term survivors of pancreatic cancer

Vinod P. Balachandran^{1,2,3}, Marta Luksza⁴, Julia N. Zhao^{1,2,3}, Vladimir Makarov^{5,6}, John Alec Moral^{1,2,3}, Romain Remark⁷, Brian Herbst², Gokce Askan^{2,8}, Umesh Bhanot⁸, Yasin Senbabaoglu⁹, Daniel K. Wells¹⁰. Charles Ian Ormsby Cary¹⁰, Olivera Grbovic-Huezo², Marc Attiyeh^{1,2}, Benjamin Medina¹, Jennifer Zhang¹, Jennifer Loo¹, Joseph Saglimbeni², Mohsen Abu-Akeel⁹, Roberta Zappasodi⁹, Nadeem Riaz^{6,11}, Martin Smoragiewicz¹², Z. Larkin Kelleyl^{3,14}, Olca Basturk⁸, Australian Pancreatic Cancer Genome Initiative⁸, Mithat Gönen¹⁵, Arnold J. Levine⁴, Ptert J. Allen^{1,2}, Douglas T. Fearon^{13,14}, Miriam Merad⁷, Sacha Gnjatic⁷, Christine A. Iacobuzio-Donahue^{2,5,8}, Jedd D. Wolchok^{3,0,14,18}, Ronald P. DeMatteo^{1,2}, Timothy A. Chan^{3,5,6,11}, Benjamin D. Greenbaum¹⁹, Taha Merghoub^{3,0,18}8 & Steven D. Leach^{1,2,5,20}8





Hot topics - microbiome

RESEARCH BRIEF

The Pancreatic Cancer Microbiome Promotes Oncogenesis by Induction of Innate and Adaptive Immune Suppression

Smruti Pushalkar¹, Mautin Hundeyin², Donnele Daley², Constantinos P, Zambirinis³, Emma Kurz², Ankita Mishra², Navyatha Mohan³, Berk Aykut², Mykhaylo Usyk¹, Luisana E. Torres², Gregor Werbar², Kevin Zhan², Yuqi Guo, Joinho Li¹, Neha Akkad², Sarah Lal², Benjamin Wadowsk², Johana Gutierrez², Juan Andres Kochen Ross², Jeremy W. Herzog³, Brian Diskin², Alejandro Torres-Hernandez², Josh Leinwand⁴, Wei Wang⁴, Pardeep S. Taunk², Shivraj Savadkar², Malvin Janal⁴, Anjana Saxena³, Xin Li¹, Deirdre Cohen⁶, R. Balfour Sartor^{2,1}, Deepak Saxena¹⁴, and George Miller^{2,9}

Tumor Microbiome Diversity and Composition Influence Pancreatic Cancer Outcomes

Erick Riquelme,^{1,2,18} Yu Zhang,^{1,18} Liangliang Zhang,^{3,4} Maria Montiel,¹ Michelle Zoltan,¹ Wenli Dong,³ Pompeyo Quesada,¹ Ismet Sahin,⁶ Vidhi Chandra,¹ Anthony San Lucas,⁶ Paul Scheet,⁶ Hanwen Xu,¹ Samir M, Hanash,^{1,7} Lei Feng,³ Jared K. Burks,⁸ Kim-Anh Do,³ Christine B. Peterson,³ Deboran Nejman,⁹ Ching-Wei D. Tzeng,¹⁰ Michael P. Kim,¹⁰ Cynthia L. Sears,¹¹ Nadim Ajami,¹² Joseph Petrosino,¹² Laura D. Wood,¹³ Anirban Maitra,¹⁴ Ravid Straussman,⁶ Matthew Katz,¹⁰ James Robert White,¹⁵ Robert Jenq,⁴ Jennifer Wargo,^{4,10} and Florencia McAllister^{1,16,17,19,*}

https://doi.org/10.1038/s41586-019-1608-2

Bacterial ablation protects against PDAC, associated with reduced MDSCs, M1-like TAM polarisation, increased TH1 CD4 T cells, and CTL activation. The PDA microbiome generates a tolerogenic immune program by activating select TLRs in monocytes?

The gut microbiome modulates the PDAC tumor microbiome landscape. A high diversity PDAC tumoral microbiome signature predicts long- term survival, with fecal microbial transplants able to modulate tumour growth and the immune TME.

LETTER

The fungal mycobiome promotes pancreatic oncogenesis via activation of MBL

Berk Aykut^{1,7}, Smruti Pushalkar^{2,7}, Ruonan Chen¹, Qianhao Li², Raquel Abengozar¹, Jacqueline I. Kim¹, Sorin A. Shadaloey¹, Dongling Wu¹, Pamela Preiss¹, Narendra Verma², Yuqi Guo², Anjana Saxena^{4,5}, Mridula Vardhan², Brian Diskin¹, Wei Wang¹, Joshua Leinwand¹, Emma Kurz¹, Juan A. Kochen Rossi¹, Mautin Hundeyin¹, Constantinos Zambrinis¹, Xin Li², Deepak Saxena^{1,2,8} & George Miller^{1,6,8} Antifungals limited PDAC progression and improved chemo efficacy. A glycan on the wall of a yeast prevalent in mouse and human PDAC is recognized by mannose binding lectin which activates C3 and the complement cascade?

Article

Fungal mycobiome drives IL-33 secretion and type 2 immunity in pancreatic cancer

Aftab Alam,¹ Eric Levanduski,¹ Parker Denz,¹ Helena Solleiro Villavicencio,^{1,13} Maulasri Bhatta,¹ Lamees Alhorebi, Yali Zhang,² Eduardo Cortes Gomez,² Brian Morreale,¹ Sharon Senchanthisai,¹ Jun Li,³ Steven G. Turowski,⁴ Sandra Sexton,⁵ Sheila Jani Sait,⁶ Prashant K. Singh,⁷ Jianmin Wang,² Anirban Maitra,⁹ Pawel Kalinski,^{1,4} Ronald A. DePinho,¹⁰ Huamin Wang,¹¹ Wenting Liao,¹² Scott I. Abrams,¹ Brahm H. Segal,⁹ and Prasenjit Dey^{1,14,4}





Hot topics - radiation

• Can figuring out what's going on in the TME in terms of immune landscape help to improve utility of RT?

Cancer Research

Check for

JEM Journal of Experimental Medicine

ARTICLE

CCR2/CCR5 inhibitor permits the radiation-induced effector T cell infiltration in pancreatic adenocarcinoma

Jianxin Wang^{1,4,5+}[®], May Tun Saung^{1,4,5+}[®], Keyu Li^{3,4,5}[®], Juan Fu^{1,4,5}[®], Kenji Fujiwara^{1,4,5}[®], Nan Niu^{1,4,5}[®], Stephen Muth^{1,4,6}[®], Junk Wang^{1,4,5}[®], Na xu^{2,2,4,5}[®], Naa Na Xu^{2,4,5}[®], Naa Na Xu^{4,5,5}[®], Naa Na Xu^{4,}

Tumor Biology and Immunology

Radiotherapy and CD40 Activation Separately Augment Immunity to Checkpoint Blockade in Cancer

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CLINICAL CANCER RESEARCH | TRANSLATIONAL CANCER MECHANISMS AND THERAPY

Modulation of the Human Pancreatic Ductal Adenocarcinoma Immune Microenvironment by Stereotactic Body Radiotherapy



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LETTER

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Radiation and dual checkpoint blockade activate non-redundant immune mechanisms in cancer

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Hot topics – vaccines

- Therapeutic cancer vaccines have historically been unsuccessful; however, recent developments have reignited interest.
- Mutant KRAS epitopes can be presented on multiple human HLA alleles and can elicit antitumor T cell responses, suggesting T cell recognition of mKRAS is possible.
- Analysis of blood from healthy donors with diverse HLA haplotypes revealed that mutant KRAS-specific T cells could be readily expanded.
- Given the early emergence of KRAS mutations, mKRAS-specific T cells may acquire tolerance?
- The development of better tools for candidate neoantigen prioritization, improved vaccine delivery platforms, and innate immune adjuvants or microbial products to engage TLR signalling and support DC activation/function could drive development?

IMMUNOTHERAPY

Bispecific antibodies targeting mutant RAS neoantigens

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Where are we going?

- Probably, more and more combos?
- Combination approaches to target tumour cells, CAFs/ECM, immune system, but also the myeloid cell mediated resistance to maximize response.

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RESEARCH

- Cancer vaccines????
- Adoptive T cell therapies????
- ???



Ongoing IO Trials

Strategy Molecular target		Cell targets	Active clinical trial(s)		
Removing short-lived immunosuppressive	CCR2	Infiltrating monocytic immunosuppressive cells	NCT03496662: BMS-813160 with nivolumab and Gem/Abraxane in borderline resectable and locally advanced PDAC		
myeloid cells	CXCR2	Infiltrating granulocytic immunosuppressive cells	NCT04477343: SX-682 with nivolumab as a maintenance therapy in metastatic PDAC		
Reprogramming DCs to better prime tumor-specific T cell responses	FLT3L and CD40 (agonism)	Mobilization of conventional DCs from bone marrow and activation in the TME	NCT04536077: Immunologic effects of CDX-301 and CDX-1140 in resectable PDAC NCT03329950: A study of CDX-1140 (CD40) as monotherapy or in combination in patients with advanced malignancies		
Reprogramming PDAC-associated fibroblasts	Vitamin D receptor	PDAC-associated fibroblasts	NCT03520790: Paricalcitol plus Gem/Abraxane in previously- untreated metastatic PDAC NCT02754726: Phase II Pilot of Paricalcitol + Gem/Abraxane + Nivolumab + Cisplatin in first-line metastatic PDAC		
Blocking immunosuppressive TGF β signaling	Angiotensin II receptor	PDAC-associated fibroblasts	NCT03563248: Losartan and nivolumab in combination with FOLFIRINOX and SBRT in localized PDAC.		
Blocking immunosuppressive TGFβ signaling	τgfβ	PDAC-associated fibroblasts	NCT04390763: NIS793 (+/- spartalizumab) in combination Gem/Abraxane in first-line metastatic PDAC		
Blocking immunosuppressive IL-1β signaling	IL-1β	PDAC-associated fibroblasts	NCT04581343: Phase 1B study of canakinumab, spartalizumab, and Gem/Abraxane in metastatic PDAC		





Vaccine Trials in PDAC

Vaccine candidate	Molecular targets	Mechanism	Phase	Trial design	Population	Primary endpoints
mRNA-5671/V941	KRASG12D, KRASG12V, KRASG13D, KRASG12C	mRNA against KRAS elicits T cell response	1	Two-arm trial: drug alone and drug plus pembrolizumab	Basket trial with KRAS- mutated PDAC, NSCLC, or CRC	Safety
ELI-002	KRASG12D, KRASG12R	Lipid-conjugated immune-stimulatory oligo + mix of lipid-conjugated peptide-based antigens	2	90 patients with PDAC randomized to vaccine or observation	Basket trial with KRAS- mutated PDAC, CRC, NSCLC, OvCa, CCa.	relapse-free survival
KRAS peptide vaccine + poly-ICLC		Targeted long peptide vaccine elicits immune response against mutant KRAS	1	Single-arm, sequential assignment of individuals	Individuals with high risk of PDAC by family history or germline mutation	Safety, IFN-producing mutant KRAS-specific CD4+ & CD8+ T cells
KRAS peptide vaccine plus poly-ICLC	KRASG12C, KRASG12V, KRASG12D, KRASG12A, KRASG13D, KRASG12R	Targeted vaccine elicits immune response against mKRAS, enhanced with ICB	1	Single-arm study of vaccine candidate + anti-PD-1 + anti- CTLA-4	Resected PDAC patients after neoadjuvant and/or adjuvant chemo and/or RT	Safety, IFN-producing mutant KRAS-specific CD8+ & CD4+ T cells
Neoantigen peptide vaccine plus poly-ICLC	Prioritized neoantigens and personalized mesothelin epitopes	Neoantigen peptide vaccine will generate neoantigen-specific CD4 & CD8 T cell responses	1	Single-arm study of PDAC patients	Resected PDAC patients after adjuvant chemotherapy	Safety
RO7198457 (mRNA-based personalized tumor vaccine)	Tumor-associated antigens	APCs take up vaccine and express antigens, leading to CTL and memory responses	1	Single-arm study in resected PDAC + vaccine + FOLFIRINOX + atezolizumab.	Resected PDAC patients	Safety
Synthetic personalized tumor-associated peptide vaccine therapy	Tumor-associated antigens	APCs take up vaccine and express antigens, leading to CTL and memory responses	I	Single-arm study of imiquimod (TLR7 agonist), pembrolizumab + vaccine	Patients with advanced PDAC or CRC	Demonstrate feasibility & safety of vaccine + pembrolizumab
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Where should we be going?

- Better models (cell of origin!)
 - Do we need germ-free facilities???
- Consideration of tumour genetics?
 - e.g. mutations linked to intrinsic resistance to ICB —
- Predictive biomarkers (e.g. NLR)
- Spatial analysis transcriptomic, multiplex IF
 - Can we target subsets of cells or aberrant signalling, to 'normalize' rather than wiping out whole cell populations
- Intravital imaging of transient interactions

Cell Reports

Tumor-associated macrophage heterogeneity is driven by tissue territories in breast cancer

Graphical abstract MMTV-PyMT CX3CR1

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Article

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In brief

The origin of tumor-associated macrophage (TAM) heterogeneity is unclear. Laviron et al. show that TAM diversity is driven by the various tissue territories existing prior to tumor apparition and by the state of tumor malignancy. This provides a definition of TAM heterogeneity according to their spatial distribution in situ.



Highlights

• TAMs differentiate according to their localization in situ

• TAM heterogeneity is associated with resident TAM diversity prior to tumor development

Orthotopic tumor models negate TAM diversity

Similar heterogeneity is found in human breast TAMs





Where should we be going?

- Pancreatitis why is the relative risk of PDAC not much higher?
- Investigation of cachexia and other systemic changes
 - Interplay between tumor and host 'normal' cells can have effects in the tumour and systemically. In turn, systemic signals may impact tumour biology.
 - Cancer-associated cachexia likely depends on specific tumour-related mechanisms including proinflammatory signals.
 - Does altered systemic inflammation/metabolism affect the tumour immune microenvironment?

Cancer Cell

CellPress

Article

Exercise-induced engagement of the IL-15/IL-15R α axis promotes anti-tumor immunity in pancreatic cancer

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Where should we be going?

- Scheduling / priming studies?
 - Priming the immune system by targeting suppressive cells prior to chemo + ICB?
 - Intermittent dosing could allow "normalised" myeloid cells to repopulate tumours and support anti-tumour immune responses (and manage tox?)
- Personalized IO strategies for individual patients or patient subgroups
- Better trials (not set up to fail!)







Conclusions

- There's a lot we don't know
- The immune TME appears, on the whole, to be tumour-promoting.
 - Is is too late to re-educate the tumour-promoting immune cells in PDAC patients? Can we only target it in terms of immunosuppression?
- There are many potential ways to transform the PDAC TME from "cold" to "hot" but we don't know which might be effective or in what combination
 - Targeting Myeloid cells / Targeting CAFs and/or ECM / Anti-CD47 to re-activate "eat me" signalling??
 / Priming innate immunity via cGAS-STING, TLRs, etc Good or bad idea? / CD40 agonists to restore cDC1 function / novel ICB targets: TIGIT, TIM3, VISTA / Cancer vaccines, Can mKRAS be harnessed as a tumour antigen? / Adoptive T cell therapies?
- Personalized immunotherapy strategies might be appropriate for individual patients or subgroups of patients ???





Points for discussion

- What are we genuinely confident that we know?!
- What don't we know that we need to?
- Can we address this? What do we need for that to happen? (tools, tech, teamwork?)
- Is this something we do well as a UK community?
 - Should we be doing this??
- Can we work better together?



